

# Optimization of RGD-modified Nano-liposomes Encapsulating Eptifibatide

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**Background:** Eptifibatide (Integrilin) is an intravenous (IV) peptide drug that selectively inhibits ligand binding to the platelet GP IIb/IIIa receptor. It is an efficient peptide drug, however has a short half-life. Therefore, antithrombotic agents like eptifibatide are required to become improved with a protected and targeted delivery system such as using nano-liposomes to the site of thrombus.

**Objectives:** The goal in the present report was to optimize encapsulation efficiency of the eptifibatide into Arg-Gly-Asp (RGD)-modified nano-liposomes (RMNL). As well, it was intended to evaluate the effect of sodium lauryl sulfate (SLS) on drug release.

**Materials and Methods:** The effect of five independent variables including number of freeze/thawing cycles, concentration of eptifibatide, 1,2-distearoyl-*sn*-glycero-3-phosphocholine (DSPC), cholesterol, and dipalmitoyl-GRGDSPA peptide on drug entrapment efficiency (DEE) was investigated using response surface methodology (RSM). The effect of different concentrations of SLS on encapsulation and drug release from RMNL was also investigated. The size and morphology of RMNL were characterized using transmission electron microscopy (TEM).

**Results:** The maximum DEE (38%) was obtained with 7 freeze/thawing cycles, 3.65  $\mu$ mol eptifibatide, 7 mM DSPC, 3 mM cholesterol, and 1 mM dipalmitoyl-GRGDSPA peptide. SLS has significantly increased the drug release from RMNL, although its effect on encapsulation efficiency was not significant.

**Conclusions:** The optimization of the formulations for valuable and expensive peptide drugs is essential to have the maximum encapsulation efficiency and the minimum experiments.

**Keywords:** Eptifibatide; RGD-modified nano-liposomes (RMNL); RMNL encapsulated eptifibatide

## 1. Background

Atherosclerosis is among the major causes of the cardiovascular disease, which results from platelet aggregation, plaque formation, ending to the occlusion of the blood vessels (1). There are chemical, protein, and peptide drugs such as eptifibatide for treating the atherosclerotic plaques efficiently. Eptifibatide (Integrilin) is an intravenous (IV) drug; a cyclic heptapeptide containing 6 amino acids and 1 mercaptopropionyl (des-amino cysteinyl) residue with an inter-chain disulfide bridge between the cysteine amide and the mercaptopropionyl moieties that selectively

inhibits ligand binding to the platelet GP IIb/IIIa receptor and rapidly dissociate from its receptor (2, 3). Eptifibatide is an efficient non immunogenic drug, however has a short half-life due to rapid inactivation and elimination by renal filtration, enzymatic degradation, and accumulation in non-targeted tissues (2). Therefore, the efficiency of the antithrombotic agents such as eptifibatide could be improved through protection and targeted delivery using nano-carriers to the site of thrombus (4).

Liposomes are the most commonly used nano-carriers for drug delivery in cardiovascular diseases (5, 6).

Nano-liposomes have numerous advantages, such as, biocompatibility, low immunogenicity, protection of the cargo against enzymatic degradation, in addition to the easy surface modification for the targeted delivery (7). Activated platelets are one of the most important targets for the targeted thrombolytic therapy. Following to the vascular injury platelet activation leads to the clot formation and restenosis (8).

In several studies, it has been shown that surface modified liposomes by fibrinogen-mimetic RGD motif achieve an affinity toward activated platelets and may be useful for the targeted delivery of thrombolytic agents (8-12). Vaidya *et al.* (2011) have developed RGD modified liposome for targeted delivery of streptokinase toward activated platelets (9).

## 2. Objectives

The main objective of this study was to develop eptifibatide encapsulation in the RGD-modified nano-liposomes (RMNL) by optimizing drug entrapment efficiency (DEE) into RMNL utilizing response surface methodology (RSM). Central composite design (CCD) was developed using Design Expert Software (DX-7, State-Ease Inc., Version 7.0.0) to evaluate the effect of the number of freeze/thaw cycles, and concentration of eptifibatide, 1, 2-distearoyl-*sn*-glycero-3-phosphocholine (DSPC), cholesterol, and dipalmitoyl-GRGDSPA peptide on the DEE of eptifibatide into RMNL. The size and morphology of the prepared liposomes were characterized utilizing transmission electron microscopy (TEM). Drug release was investigated using dialysis in both simulated body fluids (SBF) and SBF containing 0.5% sodium lauryl sulfate (SLS). SLS was incorporated in RMNL and its influence on DEE and drug release was evaluated.

## 3. Materials and Methods

### 3.1. Materials

Dimethylformamide, N-methyl pyrrolidone, and other solvents were purchased from Merck (Germany). 9-fluorenylmethyloxycarbonyl (Fmoc)-Lys (Fmoc)-OH was obtained from AAPPTEC (USA). Rink Amide MBHA resin and other protected amino acids were obtained from GLS China. 1, 2-distearoyl-*sn*-glycero-3-phosphocholine (DSPC) was purchased from Avanti Polar Lipids (USA), and cholesterol from Sigma Aldrich (USA). Eptifibatide was provided by the School of Pharmacy, Shahid Beheshti University of Medical Science (Tehran, Iran). All materials were used without any further purification.

### 3.2. Dipalmitoyl-Peptide Synthesis

Dipalmitoyl-GRGDSPA peptide conjugate was constructed from a GRGDSPA peptide head group, a KGG spacer and 2 palmitic acid (pal) anchors. This construct was synthesized manually by solid-phase peptide synthesis on Rink Amide MBHA resin using standard 9-fluoromethoxycarbonyl (Fmoc) chemistry (13). The dipalmitoyl-GRGDSPA peptide conjugate was integrated into liposomes by the pal anchors which were intercalated within the lipid bilayer. The prepared peptide was analyzed by high performance liquid chromatography (HPLC) and LC-MS spectrometry (Agilent Technology Series).

### 3.3. Liposomes Preparation and Drug Incorporation

RMNL were prepared by thin film evaporation method (14, 15). Briefly, the required amounts of DSPC, cholesterol and dipalmitoyl-GRGDSPA peptide were dissolved in 2 mL chloroform and evaporated using round-bottomed flask under low vacuum in order to form a thin lipid film. The lipid film was hydrated at 65°C in 50 mM Tris buffer at pH 7.4 for approximately 2 h. Eptifibatide solution was prepared in 50 mM Tris buffer, pH 7.4 at the desired amounts. The solution was mixed with the prepared drug free RMNL using different concentration of phospholipids and performing freeze/thawing cycles: the mixture was frozen in the liquid nitrogen (-196°C) for 5 min and thawed in water bath at 65°C and shaking at 150 rpm for 5 min. Different cycles were performed to promote the encapsulation of eptifibatide into liposomes. After optimization of the eptifibatide encapsulation, the effect of sodium lauryl sulfate (SLS) was evaluated as a surfactant on drug encapsulation and release. Different concentration of SLS solution in methanol (0.25, 0.5, 1 and 2% mol of DSPC) was added to phospholipid solution (DSPC and cholesterol and dipalmitoyl-GRGDSPA peptide dissolved in chloroform) and used for formulation of the liposomes.

### 3.4. Drug Entrapment Efficiency (DEE) Optimization

A statistical experimental design based on RSM was used to obtain optimal conditions for DEE. Based on the preliminary experiments and literature survey, five independent variables (the number of freeze/thawing cycles, in addition to concentration of the following components: eptifibatide, DSPC, cholesterol, and dipalmitoyl-GRGDSPA peptide) were selected. CCD was used with the five-variables at five-levels applying  $\frac{1}{2}$  fraction factorial and six center points resulting in 32

**Table 1.** CCD of the experiments for DEE optimization utilizing DX-7 software

Run	The number of freeze/thawing cycles	Eptifibatide ( $\mu\text{mol}$ )	DSPC (mM)	Cholesterol (mM)	Dipalmitoyl-GRGDSPA peptide (mM)	DEE (%)
1	3	1.65	6.25	3.625	0.75	0.68
2	6	2.7	7	2.75	1.5	3.31
3	3	1.65	6.25	1.875	1.25	8.62
4	8	1.65	6.25	1.875	0.75	11.16
5	8	1.65	7.75	1.875	1.25	16.29
6	3	1.65	7.75	3.625	1.25	20.25
7	6	2.7	7	2.75	0.5	3.74
8	8	3.75	7.75	1.875	0.75	24.39
9	3	3.75	7.75	1.875	1.25	25.85
10	6	0.6	7	2.75	1	27.28
11	8	1.65	7.75	3.625	0.75	27.52
12	6	2.7	5.5	2.75	1	28.14
13	6	2.7	7	1	1	29.99
14	6	2.7	7	2.75	1	27.28
15	8	3.75	6.25	1.875	1.25	32.75
16	6	2.7	7	2.75	1	35.74
17	1	2.7	7	2.75	1	33.63
18	8	3.75	7.75	3.625	1.25	33.65
19	6	2.7	7	4.5	1	33.68
20	6	2.7	8.5	2.75	1	34.64
21	6	2.7	7	2.75	1	36.95
22	3	3.75	6.25	3.625	1.25	36.23
23	6	4.8	7	2.75	1	37.79
24	8	1.65	6.25	3.625	1.25	26.53
25	3	1.65	7.75	1.875	0.75	28.01
26	6	2.7	7	2.75	1	30.81
27	8	3.75	6.25	3.625	0.75	29.74
28	3	3.75	7.75	3.625	0.75	35.23
29	11	2.7	7	2.75	1	35.59
30	6	2.7	7	2.75	1	28.98
31	6	2.7	7	2.75	1	33.17
32	3	3.75	6.25	1.875	0.75	5.71

runs (Table 1). The ranges and levels of examined independent variables are listed in (Table 2).

The percentage of DEE as response was calculated using the following equation:

$$\text{DEE (\%)} = \frac{C_{\text{initial}} - C_{\text{free}}}{C_{\text{initial}}} \times 100 \quad (1)$$

Where,  $C_{\text{free}}$  is the non-incorporated drug remained in the hydration medium following to separation of the liposomes by centrifugation at  $29000 \times g$  for 30 min. The free drug was measured by using HPLC with UV/visible detector at 220 nm. Briefly,  $4.6 \times 15$  nm reverse-phase C18 Zorbax® column (Hewlett-

**Table 2.** Experimental variables at different levels used in CCD

Independent Variables	Code	Low	Center	High	Axial (- $\alpha$ )	Axial (+ $\alpha$ )
The Number of freeze/thawing cycles	X1	3	5.5	8	1	11
Eptifibatide concentration ( $\mu\text{mol}$ )	X2	1.65	2.7	3.75	0.6	4.8
DSPC concentration (mM)	X3	6.25	7	7.75	5.5	8.5
Cholesterol concentration (mM)	X4	1.87	2.74	3.62	1	4.5
Dipalmitoyl- GRGDSPA peptide concentration (mM)	X5	0.75	1	1.25	0.5	1.5

Packard, Sigma, Germany) was used. The mobile phase was acetonitrile: water with 20:80 (V/V) ratio containing 0.1% trifluoroacetic acid.  $C_{\text{initial}}$  is the total drug used for loading drug into RMNL.

The behavior of the system was explained by the quadratic polynomial empirical model (2).

$$Y = \beta_0 + \sum_i \beta_i X_i + \sum_i \beta_{ii} X_i^2 + \sum_{i \neq j} \beta_{ij} X_i X_j \quad (2)$$

Where  $y$  is the expected response;  $\beta_0$ ,  $\beta_i$ ,  $\beta_{ii}$  and  $\beta_{ij}$  are intercept coefficient, the linear coefficient, the squared coefficient, and the interaction coefficient respectively;  $X_i$  and  $X_j$  are coded factors of independent variables (17, 18). Here  $y$  represents DEE (%) for the empirical model.

Data were evaluated for Eq.2 by Analysis of Variance (ANOVA) to assess the interaction between the independent variables and the response. The adequacy of the polynomial model was checked by the coefficient of determination ( $R^2$ ), and its statistical significance was checked by the F-test ( $p$ -value). All the independent variables were kept within the range while the response was maximized.

### 3.5. Characterization of Liposomes

The size and morphology of the prepared RMNL was analyzed by transmission electron microscopy (TEM) (Philips CM30, Netherlands). Sample were mounted on carbon coated copper grids (400 mesh, Agar Scientific, UK) and viewed without any staining. The size distribution and  $\zeta$  potential of RMNL was measured at different pH (5.5, 7.4 and 8.5) using a Zetasizer Nano ZS instrument (Malvern Instruments Ltd, Malvern, Worcestershire, UK).

### 3.6. Evaluation of the Effect of SLS Concentration on DEE and Drug Release

The influence of SLS on eptifibatide encapsulation and release was evaluated using different concentrations of SLS (0, 0.25, 0.5, 1 and 2% molL SLS.mol<sup>-1</sup> DSPC) for formulation of liposomes. Eptifibatide encapsulation was carried out based on optimum condition obtained from RSM for liposomes containing different concentration of SLS and DEE measured using HPLC analysis.

The *in vitro* release of eptifibatide from different prepared samples was examined. Briefly, liposomal samples were dispersed in 2 mL SBF and transferred into the sealed dialysis tubes (MW cut-off 12000 Da). The dialysis tubes were submerged in 10 mL SBF and SBF containing 0.5% SLS. The tubes were then incu-

bated in a thermostatic shaker (37°C, 150 rpm) for 24 h. At the pre-determined time intervals, 500  $\mu$ L of SBF containing eptifibatide released from liposomes were used for determination of the released drug concentration by HPLC. The 10 mL SBF in the recipient media was taken out and replaced with the fresh SBF at the same time intervals.

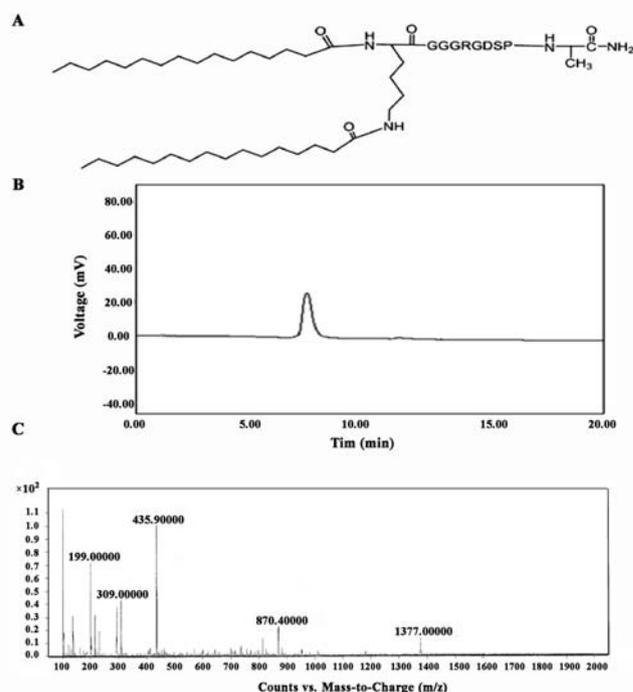
## 4. Results

### 4.1. Peptide-Dipalmitic Acid Conjugate

Dipalmitoyl-GRGDSPA peptide was synthesized by Fmoc chemistry used for surface modification of the prepared liposomes through anchoring the two palmitic acid residues to the surface of nanoliposomes. The synthesized di-palmitoyl peptide was characterized by HPLC and LC-MS analysis. Figure 1 demonstrates the structure along with the representative mass spectrum of the peptide-dipalmitic acid conjugate designed for attachment to the GPIIb-IIIa receptor. The purity of dipalmitoyl- GRGDSPA peptide conjugate was confirmed by HPLC analyses.

### 4.2. Statistical Analysis

The optimization of eptifibatide encapsulated liposomes was carried out using CCD (Table 1). The effect of the number of freeze/thaw cycles, and drug, DSPC, cholesterol and dipalmitoyl- GRGDSPA peptide con-



**Figure 1.** Dipalmitoyl- GRGDSPA peptide conjugate A: the chemical structure B: HPLC chromatogram, and C: mass spectrum

**Table 3.** ANOVA test for Response Surface Reduced Quadratic Model

Source	Sum of Squares	DF	Mean Squares	F Value	Prob > F
Model	3468.828507	14	247.7734648	13.90377	< 0.0001
A	85.72424225	1	85.72424225	4.810404	0.0425
B	463.6423275	1	463.6423275	26.01723	< 0.0001
C	220.5161087	1	220.5161087	12.37423	0.0026
D	173.066687	1	173.066687	9.711614	0.0063
E	56.67641755	1	56.67641755	3.180389	0.0924
AB	2.580477562	1	2.580477562	0.144803	0.7083
AC	198.9317198	1	198.9317198	11.16303	0.0039
AD	4.66908313	1	4.66908313	0.262005	0.6153
AE	1.508970496	1	1.508970496	0.084676	0.7746
BC	57.73953465	1	57.73953465	3.240046	0.0896
BD	77.63369473	1	77.63369473	4.356404	0.0522
BE	52.89008204	1	52.89008204	2.96792	0.1031
CE	360.5091007	1	360.5091007	20.22992	0.0003
E <sup>2</sup>	1712.740061	1	1712.740061	96.11018	< 0.0001
Residual	302.9500329	17	17.82059017		
Lack of Fit	230.5030731	12	19.20858942	1.3257	0.4010
R <sup>2</sup>	0.9197				
adj R <sup>2</sup>	0.8535				

**Table 4.** Point prediction and validation of the DEE at the optimal conditions

Confirmation Experiment (%)	Prediction	95% CI low	95% CI high
37.49	38.85	35.16	41.55

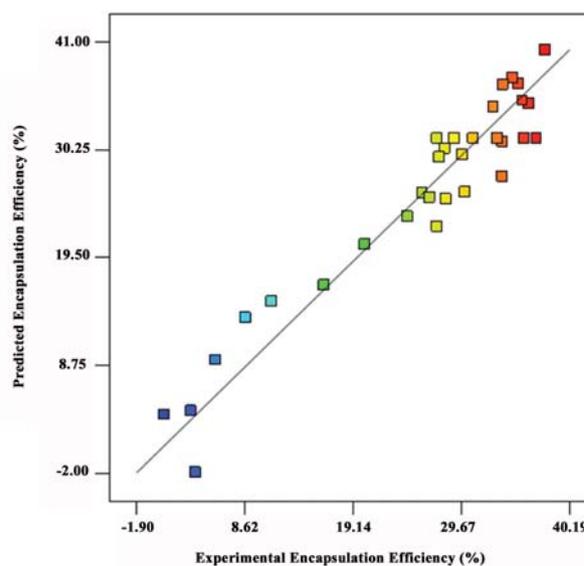
centrations on DEE were evaluated. The result of ANOVA test is presented in (Table 3) and *p*-value less than 0.05 was considered as statistically significant at 95% confidence interval.

#### 4.3. Fitting Model

A reduced quadratic model for the defined factors and their interactions are presented in equation 3.

$$R1 - 31.40 + 1.89 A + 4.40 B + 3.03 C + 2.69 D + 1.54 E - 0.40 AB - 3.53 AC + 0.54 AD - 0.31 AE - 1.90 BC + 2.02 BD - 1.82 BE - 4.75 CE - 7.56 E^2 \quad (3)$$

Where A, denotes the freeze/thawing cycle number, B, C, D, and E are the concentrations of drug, DSPC, cholesterol and dipalmitoyl-GRGDSPA peptide respectively. The relatively high R<sup>2</sup> and adj. R<sup>2</sup> values in (Table 3) indicate that model is capable of representing the system under the given experimental conditions. Also, clustering of the points around the diagonal line in (Figure 2) has confirmed a satisfactory correlation between the experimental data and the predicted values.



**Figure 2.** Predicted versus actual encapsulation efficiency (%) of the eptifibatid into RMNL obtained from the model simulation and experiments, respectively. Points are the actual amount of DEE that approximately fits with the model due to high amount of R<sup>2</sup> (0.9197)

#### 4.4. Experimental Verification

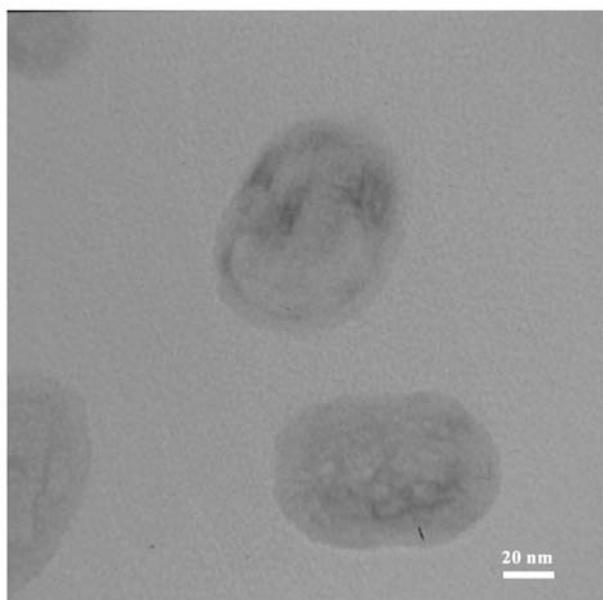
Suitability of the obtained optimum condition was validated by running an experiment (Table 4). The results of the experiment are quite close to the predicted values at a 95% of the confidence. The experimental result obtained for DEE was  $37.49 \pm 5\%$  which is within the estimated confidence interval range (35.16-41.55).

#### 4.5. Characterization of the RMNL

TEM and dynamic light scattering (DLS) analysis were used for characterizing the prepared RMNL. The results obtained from TEM analysis revealed that the size of RMNL is approximately in range of 60-90 nm (Figure 3). As well, DLS results showed that size of RMNL did not change at different pHs (Table 5). The neutral surface charge of RMNL, as shown by measurement of the  $\zeta$  potential, did not significantly change at different pH (Table 5).

#### 4.6. The effect of SLS on Eptifibatide Encapsulation Efficiency and Drug Release

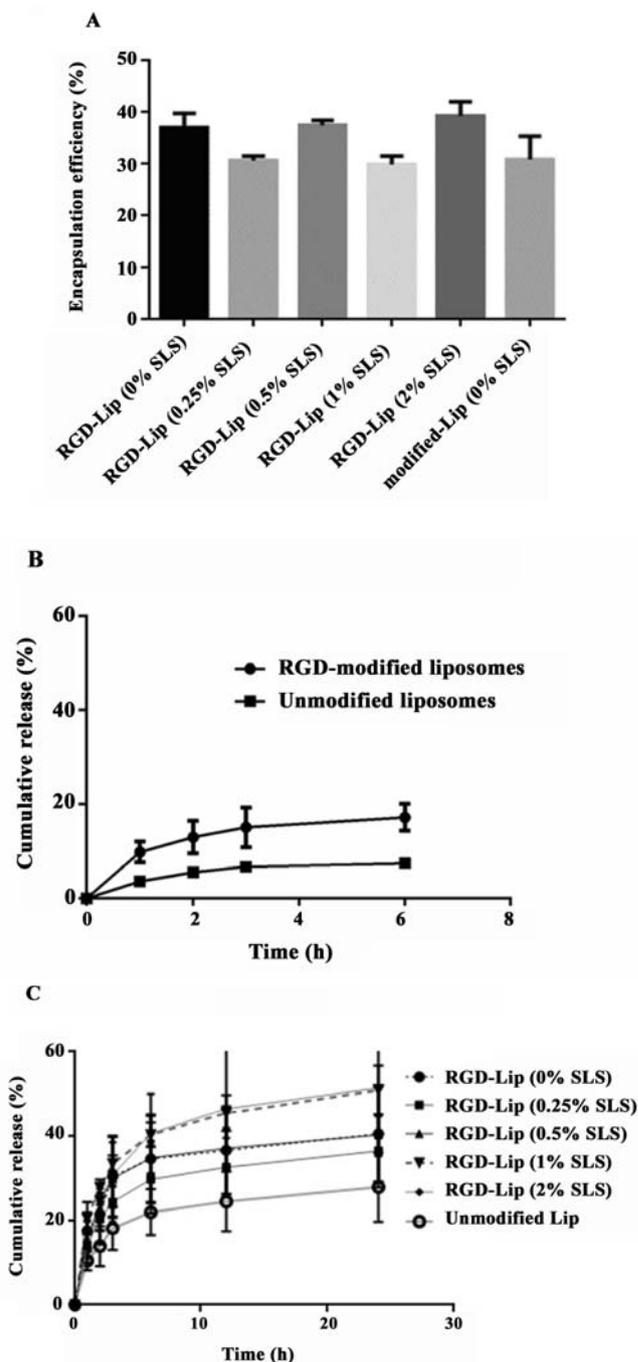
The results show that SLS effect on encapsulation efficiency of eptifibatide was not significant (Figure 4A). In addition, eptifibatide release from RMNL and UNL in SBF during 24 h was 18 and 10 % of the incorporated drug respectively (Figure 4B), whereas, drug release from RMNL and UNL was significantly enhanced in SBF containing SLS (40 and 27% of the incorporated drug during 24 h respectively, (Figure 4C).



**Figure 3.** TEM images of the phospholipid-based bilayer vesicles containing eptifibatide prepared by freeze/thawing method

**Table 5.** Size and  $\zeta$  potential measurement of RMNL at different pH

pH	Zeta (mV)	Size (nm)
5.5	-2.96	92.11
7.4	-1.44	98.21
8.5	+0.639	95.08



**Figure 4.** A: The effect of different concentration of SLS (0.25, 0.5, 1, and 2% SLS incorporated DSPE) on DEE and cumulative drug release profile of the eptifibatide-loaded liposomes at 37°C, B: only SBF, and C: SBF containing 0.5% SLS

In addition, SLS incorporated into RMNL (1 and 2% of SLS per mol of DSPC) led to a larger drug release (up to 52%) compared to UNL and RMNL without SLS (Figure 4C).

## 5. Discussion

Targeted drug delivery to the site of thrombus using fibrin specific or activated platelets specific ligands has recently become increasingly attractive (19, 20). Different peptides have been identified for targeting the various constituents of thrombus such as activated platelets (21). There are reports that have demonstrated the high affinity of the RGD peptide for GPIIb/IIIa receptor on surface of the activated platelets (10, 11, 22). In the present study, a RGD motif was used as a ligand to modify the liposome surface for targeting the activated platelet cells.

The optimization of eptifibatide encapsulation into RMNL by RSM has demonstrated a significant correlation between independent variables. Statistical analysis through application of ANOVA has further certified the suitability of the reduced Quadratic model for determination of the relationship between the variables and encapsulation efficiency (Table 4). All of the independent factors -except dipalmitoyl- GRGDSPA peptide concentration have significantly positive effect on encapsulation efficiency.

Increasing the number of freeze/thawing cycles results in the enhancement of encapsulation efficiency due to an increase in the permeability of the lipid bilayer when liposome suspensions are immersed in the liquid nitrogen. In the liquid nitrogen, ice crystals are formed across lipid bilayers (23, 24), and cause lipid fragmentation during the freezing process. During the thawing step, water channels and pores are formed as a result of ice crystals melting. Under these conditions, drug molecules are passively transferred into the inner volume of liposomes. Furthermore, according to cryopreservation experiments, ice crystals cause leakage of the incorporated drug from liposomes (25-27). With heating the liposomes above the lipid phase transition temperature ( $T_c$ ), fragmented phospholipids restore their structure and rebuild liposomes. Eptifibatide concentration also has a significant influence on the encapsulation efficiency. At higher eptifibatide concentration, the encapsulation efficiency is enhanced due to an increased entrapment of water containing hydrophilic drug. Encapsulation efficiency for hydrophilic drugs is proportional to the entrapped water volume into the inner volume of liposomes (28). Encapsulation efficiency significantly increases by

increasing the concentration of the phospholipids (DSPC) due to the enhanced number of liposomes and consequently, an increased total internal volume of the liposomes (27). The significant effect of the cholesterol on encapsulation efficiency may be related to the interaction between cholesterol and eptifibatide.

The results of drug release have confirmed the high stability of the prepared RMNL causes low drug release, which is due to acyl chain length of phospholipid (DSPC) incorporated into RMNLs. This, in turn, is directly proportional to the high transition temperature ( $T_c$ ) of DSPC ( $T_c=55^\circ\text{C}$ ) (29, 30). The increase in the drug release in the presence of SLS is due to the partial destabilization of RMNLs (31). Additionally, the high stability of the prepared liposomes by DSPC is suitable for *in vivo* application. The  $T_c$  is above body temperature; therefore reduces liposomal permeability and rate of leakage in the plasma at  $37^\circ\text{C}$  (29, 32).

At the end of this study, the critical parameters for achieving an optimal encapsulation of eptifibatide into RMNL were identified; an attractive candidate for further *in vitro* and *in vivo* studies. Excepting dipalmitoyl-GRGDSPA peptide concentration, all other factors had significantly positive effect on encapsulation efficiency. The maximum DEE (38%) was obtained with 7 cycles of freeze and thaw,  $3.65\ \mu\text{mol}$  eptifibatide, 7 mM DSPC, 3 mM cholesterol and 1 mM dipalmitoyl-GRGDSPA peptide, respectively. Our findings has provided essential information for the development of RMNL formulations in order to protect and enhance antithrombotic drugs efficacy.

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